

# Supplementary Material

## Formation of Multicolor Nanogels Based on Cationic Polyfluorenes and Poly(methyl vinyl ether-alt-maleic monoethyl ester): Potential Use as pH-responsive Fluorescent Drug Carriers

Marta Rubio-Camacho <sup>1</sup>, María José Martínez-Tomé <sup>1,\*</sup>, Amalia Mira <sup>1</sup>, Ricardo Mallavia <sup>1</sup> and C. Reyes Mateo <sup>1,\*</sup>

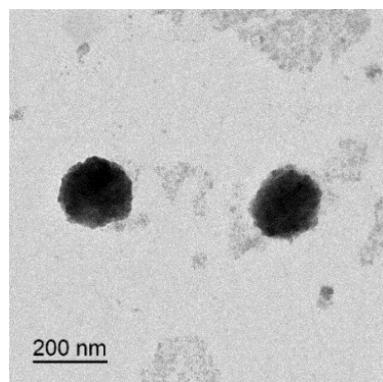
<sup>1</sup> Instituto de Investigación Desarrollo e Innovación en Biotecnología Sanitaria de Elche (IDiBE), Universidad Miguel Hernández de Elche (UMH), 03202 Elche, Alicante, Spain

**Keywords:** Nanogels; PMVEMA; fluorescent conjugated polymers; doxorubicin; nanoparticles; drug-delivery; bioimaging.

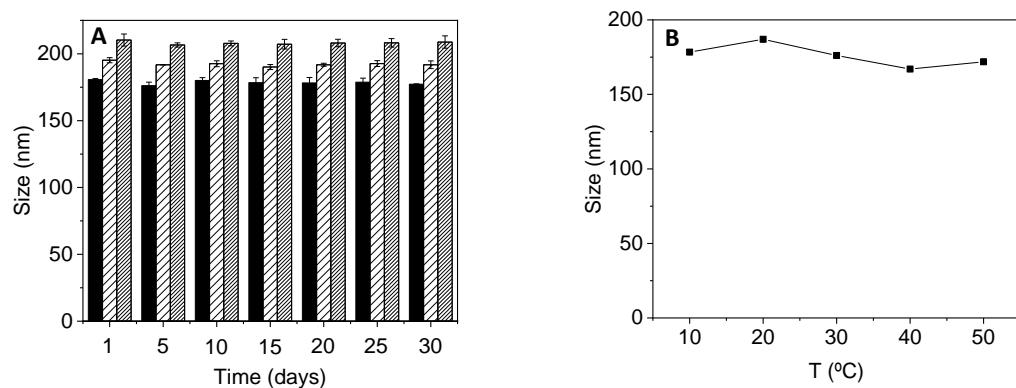
\* Correspondence: [mj.martinez@umh.es](mailto:mj.martinez@umh.es); [rmateo@umh.es](mailto:rmateo@umh.es)

**Tabla S1.** Effect of alkalinisation and subsequent acidification on the hydrodynamic diameter (d) and polydispersity index (PDI) of green and red-emitting NPs.

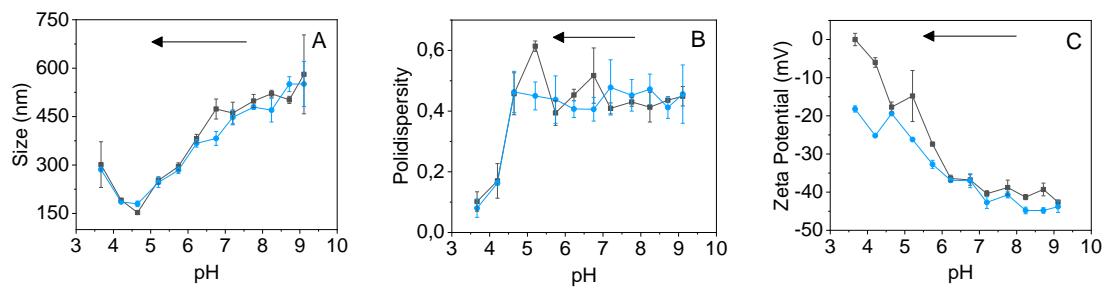
	medium	d ± sd (nm)	PDI
Green NPs	water	172.0 ± 4.2	0.12 ± 0.03
	pH 7	428.1 ± 28.6	0.25 ± 0.02
	pH 4	206.4 ± 6.0	0.13 ± 0.03
Red NPs	water	194.6 ± 4.0	0.13 ± 0.02
	pH 7	708.3 ± 11.4	0.27 ± 0.07
	pH 4	245.1 ± 8.5	0.18 ± 0.08



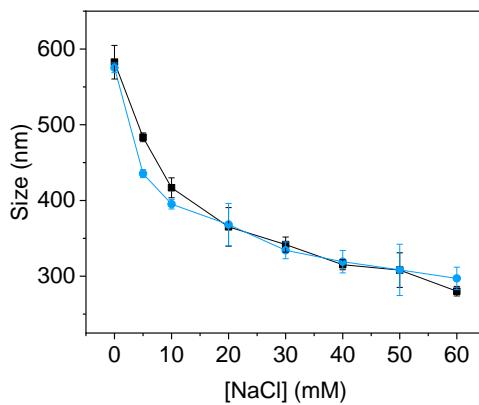
**Figure S1.** Representative images of transmission electron microscopy (TEM) of PMVEMA-Es NPs



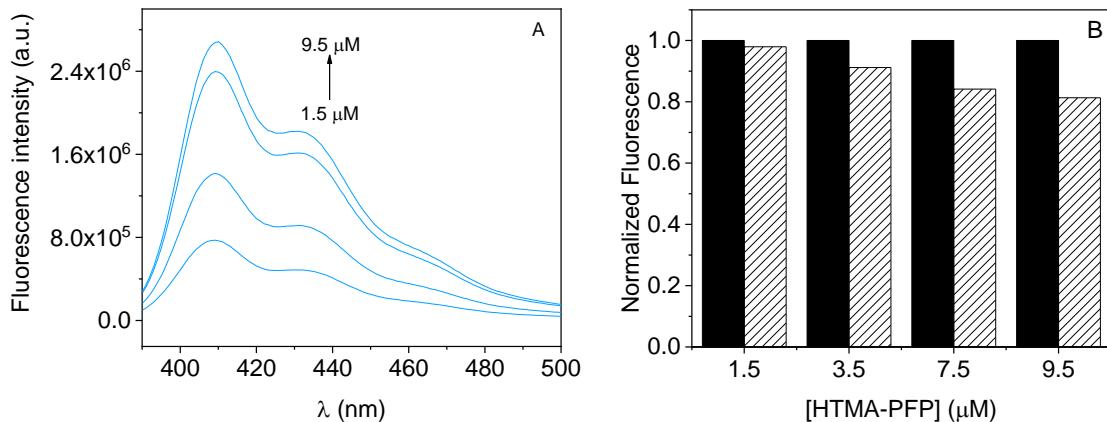
**Figure S2.** (A) Bar diagrams showing the hydrodynamic diameters of PMVEMA-Es NPs (0.03 M) as a function of time (1-30 days), in MilliQ water (black), glucose 0.2 M (striped) and sacarose 0.2 M (full striped). Note that the slight increase in size observed after addition may be related to the change in viscosity and refractive index of the medium, rather than a real increase in the size of the nanoparticles. (B) Effect of temperature (10-50°C) on the hydrodynamic diameter of NPs (0.03 M) in MilliQ water.



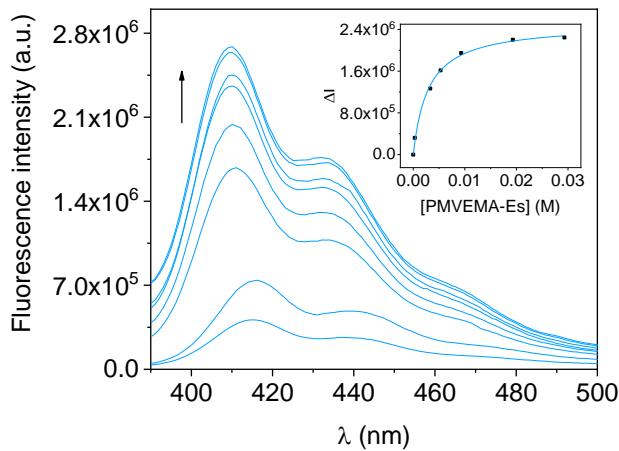
**Figure S3.** Reversible swelling–shrinking behaviour of PMEVEMA-Es NPs in absence (black) and in presence (blue) of HTMA-PFP, as the pH decreased from 9 to 3.



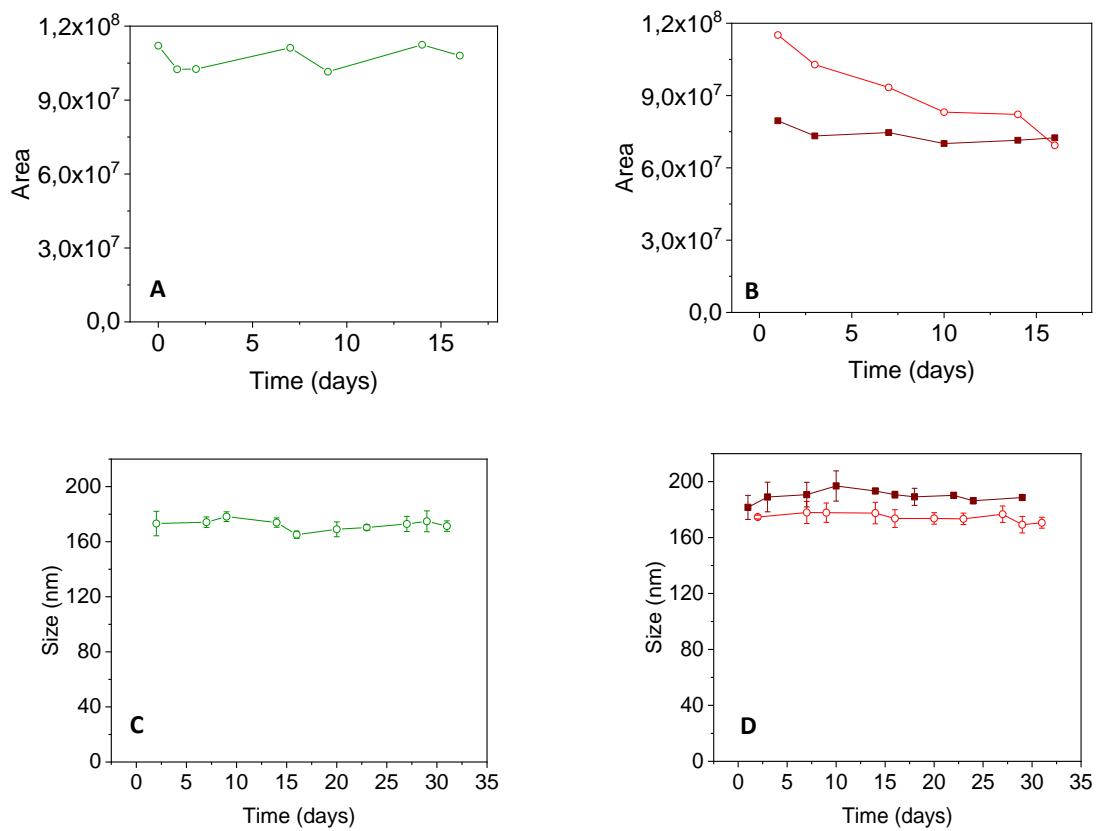
**Figure S4.** Effect of NaCl, up to 60 mM, on the hydrodynamic diameter of PMVEMA-Es NPs (0.03 M) in absence (black) and in presence (blue) of HTMA-PFP (1.5 μM).



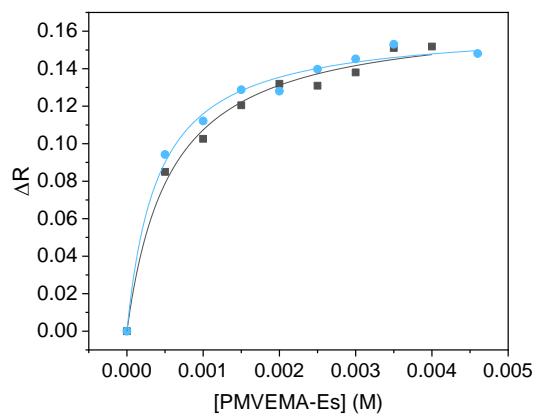
**Figure S5.** (A) Fluorescence emission spectra of HTMA-PFP in PMVEMA-Es NPs (0.03 M) at different concentrations of polyfluorene (1.5, 3.5, 7.5 and 9.5 μM). (B) Normalized fluorescence intensity of HTMA-PFP in PMVEMA-Es NPs (0.03 M) at different concentrations (1.5, 3.5, 7.5 and 9.5 μM) recorded the first (black bar) and the fifth day (strip bar) after preparation.



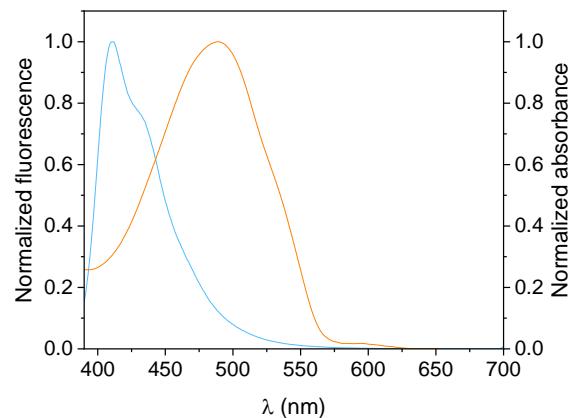
**Figure S6.** Fluorescence emission spectra of HTMA-PFP (1.5  $\mu\text{M}$ ) in MilliQ water with increasing concentrations of NPs (0-0.03 M). Inset: Changes in the fluorescence intensity of the spectrum as a function of PMVEMA-Es concentration.



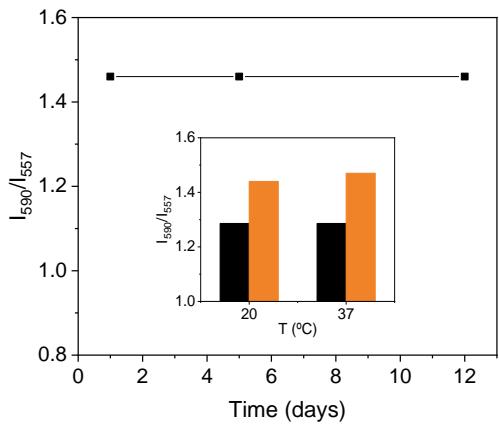
**Figure S7.** Effect of time storage on the area of the fluorescence emission spectrum (A-B) and hydrodynamic diameter (C-D) of green (A, C) and red NPs (B, D). Open circles and filled squares indicate the addition of polyelectrlyote after and during the preparation of PMVEMA-Es NPs, respectively.



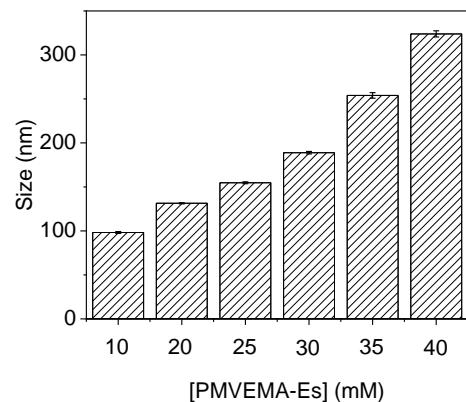
**Figure S8.** Changes in the ratio of emission peaks,  $\Delta R = (I_{590}/I_{557}) - (I_{590}/I_{557})_{\text{water}}$ , of DOX in water at increasing amounts of NPs in presence (blue circles) and in absence (black squares) of HTMA-PFP.



**Figure S9.** Overlap between the fluorescence emission spectrum of HTMA-PFP (blue) and the absorption spectrum of DOX (orange) in MilliQ water.



**Figure S10.** Effect of storage time on the ratio of the intensities of DOX emission peaks,  $R=I_{590}/I_{557}$ , inserted in the blue-emitting NPs. Inset: Effect of temperature on  $R$ , for DOX inserted in fluorescent NPs (orange bars) or free in water (black bars).  $\lambda_{\text{exc}}=380$  nm.



**Figure S11.** Hydrodynamic diameters, measured by DLS, of NPs prepared in MilliQ water, as a function of PMVEMA-Es concentration.